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(54) Title: ALPHAVIRUS EXPRESSION VECTOR (57) Abstract The present invention is directed to alphavirus expression vectors comprising at least part of an alphavirus genome and heterologous RNA inserted downstream of an alphavirus base sequence having translation enhancing activity. Such vectors can be used to achieve enhanced levels of expression of DNA or cDNA coding for a desired product and being complementary to said heterologous RNA after introduction of said vector in eukaryotic cells in cell culture or in a living body. The expression product may have therapeutical or prophylactic activity.		

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ALPHAVIRUS EXPRESSION VECTOR

The present invention is generally related to polynucleotide molecules and to their use for production of desired products after introduction thereof into human or animal cells.

5 More specifically, the present invention is directed to alphavirus expression vectors comprising at least part of an alphavirus genome and heterologous RNA inserted downstream of an alphavirus base sequence having translation enhancing activity. Such vectors can be
10 used to achieve enhanced levels of expression of DNA or cDNA coding for a desired product and being complementary to said heterologous RNA after introduction of said vector in eukaryotic cells in cell culture or in a living body.

15 The modern techniques in molecular biology have opened up many completely unforeseen possibilities for the pharmaceutical and biotechnological industry. For instance, it is today possible to clone any gene (once identified) as a DNA (or cDNA) molecule and express it
20 in prokaryotic and/or eukaryotic cells. This has facilitated the production of the corresponding proteins *in vitro* for therapeutic use and/or prophylactic use, e.g. as a vaccine. Lately the possibilities of gene expression technology have been extended also to
25 *in vivo* use in whole animals and even in human beings. Illustrative of this recent development are human gene therapy ⁽¹⁾ and genetic immunization ⁽²⁾. In order to express a foreign gene in cells *in vivo* or *in vitro*, the gene segment, or the corresponding cDNA, comprising
30 the coding sequences, is usually inserted into a so called expression vector. This vector provides for all elements that are necessary for the transcription and translation of the gene, or the cDNA thereof, within the cell. Today, there exist very efficient expression
35 vectors for bacterial and yeast cells. However, this is not the case for animal, such as mammalian, cells. This creates a large problem when a protein with mammalian

specific-modifications has to be produced and isolated for a therapeutic purpose or for prevention of disease or in cases where expression in whole organisms, e.g. the living body of an animal or human, is required.

Therefore, in very general terms there is a great demand for new expression vectors for use in animal, e.g. mammalian, cells, which vectors have

- (i) increased protein expression efficiency,
- (ii) broadened host cell specificity, and
- (iii) increased safety.

Recently, a completely new type of DNA expression vectors have been developed for use in animal, e.g. mammalian, cells. These vectors are based on the alphavirus genome.

Alphavirus is a genus belonging to the family Togaviridae having single stranded RNA genomes of positive polarity enclosed in a nucleocapsid surrounded by an envelope containing viral spike proteins.

The Alphavirus genus comprises among others the Sindbis virus, the Semliki Forest virus (SFV), the Ross River virus and Venezuelan, Western and Eastern equine encephalitis viruses, which are all closely related. In particular, the Sindbis and the Semliki Forest viruses have been widely studied and the life cycle, mode of replication, etc, of these viruses are well known and thus, need not to be specifically discussed herein.

Alphaviruses replicate very efficiently in animal cells which makes them valuable as vectors for production of protein and nucleic acids in such cells.

Expression systems based on the Sindbis virus are disclosed in US-A-5 091 309 and US-A-5 217 879. The Sindbis virus vectors of US-A-5 091 309 comprise RNA derived from Sindbis defective interfering (DI) RNA having heterologous RNA inserted therein. In US-A-5 217 879 self-replicating and self-packaging recombinant Sindbis virus RNA molecules are disclosed comprising a heterologous coding sequence and at least one Sindbis

virus junction region able to direct Sindbis virus subgenomic messenger RNA synthesis in a host cell. RNA transcripts are synthesized in vitro by transcription of Sindbis virus cDNA which has been inserted in a plasmid under control of a promoter, such as SP6.

Xiong et al., Science, Vol 243, 1989, 1188-1191 ⁽³⁾ also disclose a gene expression system based on Sindbis virus. This system is said to be efficient in a broad range of animal cells. Expression of the bacterial CAT (chloramphenicol acetyltransferase) gene in insect, avian and mammalian cells inclusive of human cells is disclosed therein.

In Bio/Technology, Volume 9, pages 1356-1361, 1991 ⁽⁴⁾, Liljeström and Garoff disclose animal cell expression vectors based on the SFV replicon. When foreign DNA coding sequences are inserted into these vectors, high amounts of foreign protein are obtained.

According to WO 92/10578, an RNA molecule is provided, which is derived from an alphavirus RNA genome and is capable of efficient infection of animal cells, which RNA molecule comprises the complete alphavirus genome regions, which are essential for replication of the said alphavirus RNA, and further comprises an exogenous RNA sequence capable of expressing its function in said host cell, said exogenous RNA sequence being inserted into a region of the RNA molecule which is non-essential to replication thereof. According to WO 92/10578 such RNA molecules can be transferred into animal cells by any means of transfection or by packaging of said RNA molecules into infectious alphavirus particles for later infection of animal cells. In both cases the transfected or infected RNA molecule will be able to replicate within the target animal cell and to express the exogenous RNA sequences inserted into said RNA molecule. Such molecules and strategies for their expression within the cell can be used as vaccines or strategies to vaccinate in order to prevent or treat

infection or cancer. In this reference, SFV has been used to illustrate alphaviruses.

The above mentioned expression vectors based on the alphavirus genome have been shown to promote a higher protein expression efficiency than earlier mammalian protein expression systems. They have also been shown to work in almost all higher eukaryotic cell types. Furthermore, they have been complemented with highly stringent safety features to prevent spreading of the virus ⁽⁵⁾. Important proteins, to be used for prevention of disease, like the HIV spike protein, have been produced with this system and such proteins have been shown to have a more native-like structure than when produced in other systems ⁽⁶⁾. Alphavirus vectors have also been used successfully for genetic immunization ⁽⁷⁾.

The present invention is directed to a significant and unforeseen improvement of the alphavirus expression vectors.

More specifically, in accordance with the present invention it has been found that there are nucleotide base sequences within the alphavirus genome, which sequences have translation enhancing activity. In comparison to the expression levels of prior known alphavirus vectors, the level of expression of a desired substance which can be obtained according to the present invention is increased about 10-fold.

Thus, the present invention is generally directed to expression of heterologous DNA in eukaryotic cells, alphavirus vectors comprising at least part of an alphavirus genome being used to achieve expression, said vectors further comprising RNA complementary to the heterologous DNA inserted essentially immediately downstream of a nucleotide base sequence having translation enhancing activity.

According to the present invention, such base sequences having translation enhancing activity (also

designated translational enhancers) comprise a 5' portion of an alphavirus capsid gene or the complete capsid gene. Suitably, this translational enhancer is endogenous to the alphavirus vector. However, exogenous translational enhancers of another alphavirus species origin might be used, at least occasionally.

Thus, the present invention is also related to a self-replicative and transcription competent recombinant alphavirus RNA molecule comprising at least part of an alphavirus RNA genome and heterologous RNA encoding a substance having biological activity located downstream of an alphavirus base sequence having translation enhancing activity, said translation enhancing base sequence being comprised of a complete alphavirus capsid gene or a 5' portion of said gene.

A suitable embodiment of the present invention is concerned with expression vectors based on the Semliki Forest virus (SFV) genome. The complete SFV capsid gene encodes 267 aminoacid residues, and, thus, comprises 801 bases.

For SFV, a translation enhancing activity has been found to reside in the first 102 bases of this capsid gene, said activity giving rise to a protein production at a level of about 85% of the wild-type capsid protein. This is an about 10-fold increase in comparison to the prior known SFV vectors.

A sequence of said gene comprising the first 81 bases also gives rise to an increased level of expression, although to a less extent.

Thus, the present SFV translational enhancer comprises at least the first 81 bases, and preferably at least the first 102 bases of the capsid gene and at most 801 bases, i.e. a base sequence corresponding to the complete capsid gene.

The sequence of the said 5' portion of the SFV capsid gene comprised of the first 102 bases, reading from 5' to 3' end, is

ATGAATTACA	TCCCTACGCA	AACGTTTTAC
GGCCGCCGGT	GGCGCCCGCG	CCCGGCCGCC
CGTCCTTGGC	CGTTGCAGGC	CACTCCGGTG

GCTCCCGTCG TC.

Modifications of this base sequence (e.g. base deletions, substitutions, and/or additions) having an essentially conserved enhancing activity are also encompassed by the present invention.

In the following illustrative Examples the occurrence of a translational enhancer has been demonstrated for SFV. In view of the homology between various alphavirus species, it can be expected that a similar mechanism of translation enhancement exists in all alphaviruses. However, as the sequence between different alphaviruses vary considerably in the 5' end of the C gene region, it is most likely that some similarities in the features of the secondary and the tertiary structure of the RNA molecule are responsible for the translation enhancing effect. This means that the exact sequences and probably also the length of the translation enhancing regions will vary between different alphaviruses.

The present invention is also related to a DNA molecule comprising DNA sequences complementary to the present recombinant alphavirus RNA molecule, which DNA molecule also may comprise a DNA sequence encoding a promoter, such as SP6, for transcribing the recombinant RNA molecule in cells and additional DNA sequences encoding traits required for plasmid growth in *E. coli*.

The present alphavirus vectors having enhanced translation capacity can be used for the same purposes as the previously known alphavirus vectors. Thus, they are likely to be very useful for production of substances having biological activities, such as proteins or polypeptides, in eukaryotic cells, especially mammalian ones, which substances can be used for biotechnical or medical purposes. For instance, the heterologous RNA may suitably encode a protein, polypeptide or peptide having therapeutic activity or prophylactic, such as immunogenic or antigenic, activity. Said eukaryotic cells, wherein expression is achieved, can be available as cell cultures or constitute a part of a living organism, such as an animal or human being. The present invention is also concerned with cells

transformed with the present recombinant RNA or DNA molecule and cell lines stably transformed with said RNA or DNA molecule.

As stated above, the alphavirus expression vectors are based on the alphavirus genome. This consists of a single-stranded RNA molecule of positive polarity. In the infected cell the 5' 2/3 of this RNA serves as a mRNA for the viral nonstructural polyprotein, which is co- and posttranslationally cleaved into four mature proteins (8, 9). These proteins form the viral polymerase complex which replicates the genomic RNA via an intermediate of negative polarity (10). This intermediate also functions as a template for the synthesis of a subgenomic RNA molecule, which is colinear with the 3' 1/3 of the viral genome. This subgenomic transcript (also called 26S mRNA) is translated into the viral structural poly-protein. Processing of this polyprotein is initiated by the amino terminal capsid (C) protein, which autocatalytically cleaves itself from the nascent polyprotein chain (11, 12, 13, 14). The remaining part of the structural polyprotein is co-translationally inserted into the membrane of the endoplasmic reticulum and the spike proteins p62 and E1 are released by signal peptidase-mediated cleavage events (15, 16, 17).

The alphavirus expression vectors are suitably constructed from a modified cDNA copy of the viral genome, from which at least part of the structural polyprotein-encoding region has been deleted and replaced with a cloning site (3, 4). Heterologous cDNA can be inserted into this site and the corresponding recombinant alphavirus genome can be produced by *in vitro* transcription. When transfected into host cells the recombinant genome is replicated in a wild-type manner since it contains both the non- structural coding region of the alphavirus and the 5' and 3' replication signals. However, instead of the virus structural proteins the subgenomic RNA now directs the synthesis of the heterologous protein. The Semliki Forest

virus (SFV) expression system ⁽⁴⁾ has also been supplemented with an *in vivo* packaging system whereby recombinant genomes can be packaged into infectious SFV-like particles following co-transfection with a
5 packaging-deficient helper genome, which provides the viral structural proteins ^(4, 5). These recombinant particles can be used to infect cells either *in vivo* or *in vitro*. The host cell range of the recombinant particles is determined by the alphavirus spike and is therefore
10 very broad. The infected cells will produce recombinant genomes and also high amounts of the heterologous proteins. However, as no virus structural proteins are encoded by the recombinant genomes, no new virus particles will be made, and hence there is no spreading of
15 virus.

Accordingly, one embodiment of the present invention is directed to a method to produce a recombinant alphavirus comprising a recombinant alphavirus RNA genome surrounded by a wild-type alphavirus coat, the said
20 RNA genome comprising the present recombinant RNA molecule, by co-transformation, such cotransfection, of cells with the said recombinant RNA genome and a helper RNA containing expression capacity of alphavirus structural proteins and comprising coding sequences for the
25 alphavirus structural proteins, *cis* acting replication signals but no encapsidation signals, incubation of the cells and collection of medium containing infectious recombinant alphavirus particles.

The present invention is also related to these co-transformed cells producing the infectious particles
30 and to the infectious particles *per se*.

As indicated above, a broad range of host cells of animal (including human) origin can be used according to the present invention. Such host cells can be
35 selected from avian, mammalian, amphibian, insect and fish cells. Illustrative of mammalian cells are human, monkey, hamster, mouse and porcine cells. Suitable

avian cells are chicken cells.

Furthermore, the present invention can be used both *in vitro* and *in vivo*. As per definition, *in vitro* means a process performed outside a living organism as
5 opposed to *in vivo* which means that a process is performed inside a living organism.

In accordance with the present invention "transformation" is intended to mean introduction in general of exogenous polynucleotides sequences into the interior
10 of a cell, eukaryotic or prokaryotic, and the exogenous polynucleotide sequence may remain in cell cytoplasm, in nucleus as extrachromosomal (episomal) or may be stably integrated into the cell genome. The mode of transformation is not crucial, but any means, known at
15 present or that may be developed in the future, can be used according to the invention.

A further embodiment of the invention is related to a method for expressing a heterologous RNA sequence encoding a biologically active substance comprising
20 infection of cells in cell culture or in an animal or human individual with the present infectious recombinant alphavirus particles.

Suitably, such expression, when achieved in an animal, will not give rise to any effect beneficial to
25 said animal, but the expression product produced in said animal can be recovered from said animal, e.g. in a body fluid, such as blood, milk or ascites.

The present invention is also related to a pharmaceutical preparations comprising the present recombinant RNA in a physiologically administrable form. Such
30 preparations may comprise the present co-transformed cells producing infectious recombinant alphavirus particles or such infectious particles per se.

In the following experimental part, the Semliki Forest virus (SFV) has been used to illustrate the
35 present invention. The earlier described SFV expression system provides three different plasmid vectors

(pSFV1-3) with a cloning site for heterologous DNA in, or immediately after, the 5' untranslated leader sequence of the subgenomic 26S RNA ⁽⁴⁾. In Example 2 the expression levels of several different heterologous proteins are compared when using the earlier SFV vectors and when using a new SFV vector in which the complete 26S RNA leader plus the complete C protein gene precedes the heterologous protein gene (=the SFVC-vector). It is evident that heterologous products are produced from SFVC-vectors at a substantially higher level than from the earlier vectors and that all C-fusion proteins have been cleaved into separate authentic products via the autoprotease activity of the C protein (Fig. 2). Quantitation shows that there is about a ten-fold difference in expression level.

In example 3 we have used E. coli LacZ (β -galactosidase) gene fusions in order to study where in the C gene the enhancing effect resides. Specifically, various portions of the 5' part of the C gene have been fused to the second codon of the LacZ gene. Expression of these recombinant constructions show that the enhancing effect is located within the 102 first bases of the C gene.

In example 4 we have analyzed whether the effect is expressed on the level of RNA replication and transcription. The production of genomic RNA and subgenomic RNA was compared in high and low producer variants of the LacZ recombinants described in example 3. No differences in RNA levels were found and hence the enhancing effect of the C gene segment must reside on the translational level. This segment most likely contains a specific RNA structure that enhances translational initiation.

Description of drawings

Fig. 1.

A. Overview of recombinant subgenomic RNA molecules. Shown are schematic drawings of the subgenomic RNA

molecules produced in cells that have been transfected with *in vitro* transcripts of the indicated plasmids. Note, that only part of the untranslated 5' leader sequence is present in the pSFV1 constructs, whereas the whole leader sequence is present in the pSFV3 constructs ⁽⁴⁾.

B. Number of C-protein amino acid codons in plasmids of the pSFVC'-series.

Fig. 2

The presence of the C gene enhances protein production.

BHK-21 cells were transfected with RNA transcripts of the following plasmids; pSFV1-p62, pSFVC-p62, pSFV1-TR, pSFVC-TR, pSFV1-Pr65^{gag}, pSFVC-Pr65^{gag}, pSFV3-LacZ, pSFVC-LacZ, pSFVmSQL and pSFVC-Pr65^{gag} (lanes 1-10, respectively). At 8 h post transfection cells were pulse-labelled with ³⁵S-methionine for 30 min (lanes 1-6) or 15 min (lanes 7-10) and chased for 15 min (lanes 1-10). Cells were lysed with NP-40 (lanes 1-6) or SDS (lanes 7-10) and aliquots corresponding to 1.6 x 10⁵ cells were either extracted with antibodies against SFV E2 (lanes 1-2), the transferrin receptor (lanes 3-4) or against Moloney Murine Leukemia virus (lanes 5-6) or mixed directly with sample buffer (lanes 7-10) and analyzed by SDS-PAGE under reducing conditions followed by fluorography.

Fig. 3

A 102 base segment from the C gene is responsible for protein expression enhancement.

BHK-21-cells were transfected with RNA transcripts of pSFV3-LacZ, six members of the SFVC'-LacZ series and pSFVC-LacZ as indicated. At 8 h post transfection cells were pulse-labelled with ³⁵S-methionine for 15 min, chased for 15 min and then lysed with SDS. Aliquots of cell lysates (corresponding to 1.2 x 10⁴ cells) were mixed with sample buffer and analyzed by SDS-PAGE under reducing conditions. The amount of β -galactosidase or β -galactosidase fusion proteins were quantitated as

described in example 2 and corrected for differences in methionine content. Shown is the relative rate of protein synthesis (expressed as percentage of SFV_{MSQL} C-protein synthesis) versus length of the variable C' part of the C'- β -galactosidase fusion proteins.

Fig. 4

Expression enhancement by the 102 base long C gene segment is not mediated through increased replication nor transcription.

Cells were transfected with RNA transcripts from pSFV_{MSQL} (lane 1), pSFVC'b7-LacZ (lane 2), pSFV3-LacZ (lane 3), pSFV1-Pr55^{gag} (lane 5) or with buffer alone (lane 4). Actinomycin D (1.0 μ g/ml) was added 2 h post transfection. At 4 h post transfection ¹⁴C-uridine was added and the cells labelled for 4 h. Total RNA was extracted and aliquots corresponding to 1.4×10^5 cells were analyzed on an agarose gel under denaturing conditions. Shown is an autoradiogram of the dried gel.

Examples

Example 1

In this we describe the construction of SFV vectors with heterologous genes that are fused either to the 26S mRNA leader, to the 3' end of the complete C gene or to 5' portions of the C gene. All constructions have been done using the complete SFV cDNA molecule in plasmid pSP6-SFV4⁽⁴⁾ which have been deposited at the European Collection of Animal Cell Culture, Porton Down, Salisbury, Wiltshire, U.K.; Accession number 91112826. In Fig. 1 we show schematically the coding regions of the various mRNA molecules that are produced from the different vector constructs. The vector constructs are:

- (i) pSFVC-p62, which contains the gene for the SFV p62 spike subunit as a wild-type fusion with the C gene. It was constructed by ligating the large SpeI/NsiI-fragment (12 kbp) from pSP6-SFV4 to the small NsiI/SpeI-fragment (2.6 kbp) of a

pSP6-SFV4 where two termination codons had been introduced at position 9866-9871 in pSP6-SFV4, in the region encoding the N-terminal part of E1 by site directed mutagenesis.

5 (ii) pSFV1-p62, which contains the gene for the SFV p62 spike subunit fused to the 5' SFV subgenome region at position 31 in the 26S mRNA leader sequence (SFV-1 vector variant) ⁽⁴⁾. This was constructed by ligating the large SpeI/NsiI-
10 fragment (11kbp) from pSP6-SFV4/ Δ C to the small NsiI/SpeI-fragment (2.6 kbp) in (i). The pSP6-SFV4/ Δ C was constructed from pSP6-SFV4 by a deletion of the C gene ⁽¹⁸⁾.

15 (iii) pSFV1-TR, which contains the human transferrin receptor gene in a fusion with the 5' SFV subgenome region at position 31 in the 26S RNA leader sequence ⁽⁴⁾.

(iv) pSFVC-TR, which contains the human transferrin receptor gene in a fusion with the 5' SFV
20 subgenome encompassing the complete 26S RNA leader sequence plus the whole C gene. The precise fusion of C and TR genes was carried out by PCR ^(19, 20). The terminal primers used for the amplification of the fused DNA were: 5'
25 CAACGGAAAAACGCAGCAGC 3' (the 5' end primer) and 5' CTTTGCTGAGTTTAAATTCACG 3' (the 3' end primer). The fusion primers used were: 5' GATCTAGCTTGATCCATCATCCACTCTTCGGACCCCTCGGG 3' (C gene fragment primer) and 5'
30 CCCGAGGGGTCCGAAGAGTGGATGATGGATCAAGCTAGATCAGC 3' (TR gene fragment primer). pSFVC-TR was constructed by ligating the 896-bp Csp45I-NdeI fusion fragment to the 10679-bp SpeI-Csp45I fragment of pSFV-C ⁽¹⁸⁾ and the 2744-bp NdeI-SpeI
35 fragment of pSFV1-TR ⁽⁴⁾. The 896-bp Csp45I-NdeI fragment of pSFVC-TR originating from the PCR reaction was checked by sequencing.

(v) pSFV1-Pr65^{gag}, which contains the gene for the Moloney Murine Leukemia virus (M-MuLV) gag precursor in fusion with the 5' SFV subgenome region at position 31 in the 26S mRNA leader sequence. This was constructed by insertion of a 1770-bp *Pst*I-*Dra*III-fragment from pNCA (Colicelli, 1988) containing the Pr65^{gag} gene, into pSFV1 via subcloning steps in pGEM-1 and pGEM-7Zf (+) (Promega). The subcloning steps introduced a 5' *Bam*HI and a 3' *Sma*I site flanking the Pr65^{gag} gene. The 5' noncoding M-MuLV sequence was deleted by replacing the 5' 177-bp *Bam*HI-*Pst*I fragment with a synthetic 135-bp *Bam*HI-*Pst*I fragment, which was made from the following two overlapping oligonucleotides; (1) 5' ATTCTGATTGGATCCAGCACCATGGGCCAGACTGTTACCACTCCCTTA AGTTTGACCTTAGGTCACTGGAAAGATGTCGAGCGGATCGCTCACAAC CAG 3' and (2) 5' GGTGGCCATTCTGCAGAGCAGAAGGTAACCCAACGTCTCTTCTTGAC ATCTACCGACTGGTTGTGAGCGATCCGCTCGACATCTTTCCAGTGACC TAA 3'.

(vi) pSFVC-Pr65^{gag}, which contains the M-MuLV gag precursor gene in a fusion with the SFV 5' subgenome region encompassing the complete 26S RNA leader plus the C gene. This plasmid was constructed by a PCR fusion of the SFV C gene to the second codon of the Pr65^{gag} gene. The fusion primers used were: 5' GGGAGTGGTAACAGTCTGGCCCCACTCTTCGGACCCCTCGGG 3' (C gene fragment primer) and 5' CCCGAGGGGTCCGAAGAGTGGGGCCAGACTGTTACCACTCCC 3' (Pr65^{gag} gene fragment primer). The terminal primers for amplification of the fused DNA were: 5' CAACGGAAAAACGCAGCAGC 3' (the 5' end primer) and 5' CAAGGCTTCCCAGGTCACG 3' (the 3' end primer). The pSFVC-Pr65^{gag} was then constructed by ligation of the 562-bp *Csp*45I-*Pst*I C-Pr65^{gag}

fusion fragment to the 10679-bp *SpeI*-*Csp45I* from pSFV-C⁽¹⁸⁾ and the 2492-bp *PstI*-*SpeI* fragment of pSFV1-Pr65^{gag}. The 562-bp *Csp45I*-*PstI* fragment of pSFVC-Pr65^{gag} originating from the PCR reaction was checked by sequencing.

(vii) pSFV3-LacZ, which contains the LacZ (β -galactosidase) gene encoding amino acid residues 2-1025 in a fusion with the 5' SFV subgenome region at position 53 in the 26S RNA after the complete untranslated leader sequence (SFV3 vector, ⁽⁴⁾).

(viii) pSFVC-LacZ, which contains the LacZ gene as above in a fusion with the SFV 5' subgenome region encompassing the complete 26S mRNA leader sequence plus the complete C gene. This was constructed by insertion of the small LacZ-containing *Bam*H1-fragment (3 kbp) from pSFV3-LacZ in the pSFVC^{BamH1}. The pSFVC^{BamH1} contains only the C protein gene of the structural SFV genome, directly followed by a *Bam*H1 site. This plasmid was constructed by ligation of the large *SpeI*-*Csp45I* fragment (10679-bp) from pSP6-SFV4 to a PCR-generated 440-bp *Csp45I*-*Bam*H1 3' end fragment of the C gene and the 874-bp *Bam*H1-*SpeI* fragment of pSFV1 (ref. Liljeström & Garoff, Biotechnology 1991). Primers used in the PCR reaction were: (1) 5' CAACGGAAAAACGCAGCAGC 3' (the 5' end primer) and (2) 5' CTCAAG GAGTCATGGATCCCACTCTTCGGACCCCTCGGG 3' (the 3' end primer).

(ix) pSFVC'-LacZ, which contains the LacZ gene in a fusion with the 5' subgenome region encompassing the complete 26S mRNA leader plus a 5' portion of the C gene. This was constructed as follows: serial deletions in the C-protein gene of pSP6-SFV4 was made by limited digestion with nuclease Bal 31, starting from the *Csp45I* site

at position 7781. The ends were blunted with Klenow fragment and T4 DNA polymerase and ligated to a mixture of *Bam*H1-linkers (8-, 10- and 12- mers; Promega, Madison, WI, USA). The large *Spe*I-*Bam*H1 fragments (approximately 10.3 kbp) from the deletion series were ligated to the small, LacZ gene containing *Bam*H1-*Spe*I fragment from pSFV3-LacZ.

(x) pSFVmSQL, which is a control plasmid and contains the complete SFV genome. It harbours mutations in the structural gene region which specifies the maturation cleavage site of p62. Because of these mutations the p62 subunit is not cleaved to E2 and E3 (21, 5).

The detailed protocols for all molecular biological techniques used are described in Molecular Cloning (22).

Example 2

The presence of the C gene enhances protein production

RNA was synthesized *in vitro* using plasmids pSFV1-p62, pSFVC-p62, pSFV1-TR, pSFVC-TR, pSFV1-Pr65^{gag}, pSFVC-Pr65^{gag}, pSFV1-LacZ, pSFVC-LacZ and pSFVmSQL as templates. These were transfected into BHK-21 cells and at 8 h post transfection the cells were metabolically labelled with ³⁵S-methionine for 15 or 30 min as indicated in the legend of Fig. 2. Fig. 2 shows the relative level of synthesis of the homologous p62-protein (lanes 1 and 2), the transferrin receptor (lanes 3 and 4), the M-MuLV gag-precursor protein (lanes 5, 6, and 10), the β -galactosidase-protein (lanes 7 and 8) when expressed alone (lanes 1, 3, 5 and 7) or in tandem with the C-protein (lanes 2, 4, 6, 8 and 10). The synthesis of the structural SFV proteins (C, p62 and E1) are shown in lane 10. Immunoprecipitations from cell lysates (lanes 1-6) or aliquots of total cell lysates (lanes 7-10) are shown.

When expressed from pSFV1-based recombinant genomes SFV p62, TR and M-MuLV gag (lane 1, 3 and 5, respectively) were all synthesized at a comparable low level. However, when these proteins were expressed as C fusion proteins, a dramatic increase in their expression levels were obtained (lanes 2, 4 and 6). Quantitations showed that the increase was approximately 10-fold for all three proteins. The autolytic cleavage that releases the C-protein from the virus structural polyprotein was apparently effective also in the artificial C-Pr65^{gag} and C-TR polyproteins; there was no sign of slow migrating polypeptides which could represent uncleaved fusion proteins (lanes 4 and 6).

In the pSFV1-based genomes only the 5' part of the 26S leader sequence is present, while the subgenome derived from pSFVC contain the whole 5'-leader. Therefore, the results indicate that the 3' part of the leader sequence or the presence of the C-sequence influence the expression level. In order to find out which region was responsible for enhancing expression we compared the level of β -galactosidase production in cells transfected with either the SFV3-LacZ genome (which contains the whole leader sequence (Fig. 2, lane 7) or the SFVC-LacZ genome (which contains the whole leader sequence plus the C gene sequence) (Fig. 2, lane 8). The latter genome directed the synthesis of an uncleaved C- β -galactosidase fusion protein (1.263 amino acids long, approximately 145 kDa). Quantitation showed that the expression of the C- β -galactosidase fusion protein was approximately 8 times higher than the expression of β -galactosidase from the SFV3-LacZ genome. This increase was as large as that observed above for SFV1 in comparison to SFVC. From these observations we conclude that the presence of C gene sequence in the expression vector has a strong positive effect on SFV-driven production. The most likely reason

for the inhibition of cleavage in the C- β -galactosidase construct is a proline residue at position +2, counted from the site cleaved by the capsid protease.

To correlate the level of protein production from an SFVC-genome to that from the wild-type-like SFV genome, we compared the amount of C-protein produced in cells transfected with *in vitro* transcripts of pSFVmsQL (Fig. 2, lane 9) to the amounts of C and gag-precursor in cells transfected with the SFVC-Pr65^{gag} RNA (Fig. 2, lane 10). The amount of C-protein at 8 h post transfection was indistinguishable in the two transfections. Furthermore, Pr65^{gag} and SFV spike proteins were produced at comparable levels, thus showing that the introduction of the C-gene into the expression vector increased the production of heterologous proteins to a level normally obtained for the structural proteins in a wild-type SFV infection.

In this example RNA transcription and transfection was done as follows: RNA transcripts of SpeI-linearized plasmids were synthesized *in vitro* as described previously ⁽⁴⁾ and transfected into BHK21 cells (American Type Culture Collection) by electroporation. In brief, 8×10^6 cells (suspended in 0.8 ml of Ca²⁺/Mg²⁺-free PBS) were mixed with *in vitro* made RNA (20 μ l of the transcription mix) in a 0.4-cm electroporation cuvette (Bio-Rad Laboratories AB, Solna, Sweden) and exposed to two consecutive pulses at 0.85 kV and 25 μ F (BioRad Gene Pulser, without the pulse controller unit). The cells were suspended in 15 ml complete BHK-medium, consisting of BHK-21 medium (Gibco) supplemented with 10 % (v/v) tryptose phosphate broth (Gibco, Madison, WI, USA), 5 % (v/v) fetal calf serum, 20 mM N-2-hydroxyethylpiperazine-N'-2-ethanesulphonic acid (HEPES) pH 7.3, plated on 3.3 cm tissue culture dishes (3 ml/dish) and incubated at 37°C.

Metabolic labelling with ³⁵S-methionine was done as

follows: At 8 h post transfection the cells were rinsed twice with PBS, 1.0 ml Eagles minimal essential medium lacking methionine and supplemented with 20 mM HEPES was added and the cells incubated for 30 min at 37°C.

The medium was removed and 3.7 MBq (100 μ Ci) 35 S-methionine in 1.0 ml of the same medium was added and the cells incubated for 15 or 30 min (pulse). The labelling medium was removed and 1.0 ml of EMEM supplemented with 20 mM HEPES was added and the cells further incubated for 15 or 30 min (chase). At the end of the chase the cells were rinsed twice with PBS and solubilized in 0.30 ml of 10 mg/ml SDS or 10 mg/ml Nonidet P40 (NP-40) lysis buffer containing 50 mM Tris-HCl, 0.15 M NaCl and 2.0 mM EDTA, final pH 7.6. Phenylmethylsulfonyl fluoride (PMSF; 0.2 mM), N-ethylmaleimide (0.1 mM) and pepstatin A (1 μ g/ml) was added to the lysis buffer before use. Cells disrupted with SDS lysis buffer were sonicated for 20 min at room temperature. Cell nuclei from NP-40 lysed cells were pelleted by centrifugation (5 min at 14.000 x g).

Immunoprecipitations were done as follows:

Transferrin receptor was precipitated with the OKT9 monoclonal anti-TR antibody (a gift from T. Ebel) the M-MuLV gag precursor with a polyclonal anti-MuLV serum (HC 185; Quality Biotechnology Incorporated) and the p62 with a monoclonal anti E2-antibody (UM 5.1; Boere et al. 1984). Aliquots of SDS-lysates of 35 S-methionine labelled cells (100 μ l) were diluted 10 times with 10 mg/ml NP-40 lysis buffer, and an excess of the appropriate antibody was added together with 40 μ l protein A-Sepharose slurry (Pharmacia, Uppsala, Sweden; 50 % w/v in 10 mM Tris-HCL, pH 7.5) and the samples were rotated end over end for 16 h at +4° C. When linking antibodies were necessary, rabbit anti-mouse immunoglobulin (Dacopatts, Glostrup, Denmark) were also added. The immune complexes were washed as described

(Wahlberg and Garoff 1989) and prepared for SDS-PAGE.

SDS-PAGE was done as follows: Washed immune complexes or samples of cell lysates were mixed with SDS-PAGE sample buffer (giving the final concentrations; 0.15 M Tris-HCL, pH 8.8, 0.19 g/ml glycerol, 31 mg/ml SDS, 3.8 mM EDTA, 0.77 mM methionine, 0.15 mg/ml bromphenol blue and 38.5 mM dithiothreitol) heated for 3 min at 95°C, cooled and alkylated with iodoacetamide (final concentration 0.10 M). Finally, the proteins were separated on 11 % acrylamide gels as described (Maizel 1971). After electrophoresis the gels were immersed in 10 % (v/v) trichloroacetic acid and 40 % (v/v) methanol, incubated for 30 min and dried. Gels containing radio-labelled material were exposed to Kodac XAR films for autoradiography.

Quantitations were done as follows: Dried acrylamide-gels that contained radio-labelled material were exposed to a Bas-III Image Plate (Fuji Photo Film Corp.). The image plate signals were digitized and the amount of radioactivity in different parts of the dried gels were analyzed using the Fuji Bio-Image analyze system Bas 2000 (Fuji Photo film Corp.).

Example 3

A 102 base segment from the 5' portion of the C gene is mediating the expression enhancement.

As a first step in elucidating the mechanism of expression enhancement we addressed the question whether the full C-gene sequence, or only a portion of it, was sufficient to promote high levels of protein production. To this end various plasmids (C'-LacZ chimeras), which contained increasing portions of the 5' part of the C-gene (C') fused to the β -galactosidase-gene were constructed (Fig. 1). These plasmids were used as templates for RNA transcription in vitro, the RNAs were transfected into BHK-21 cells and the cells were pulse-labelled for 15 min and chased for 15

min as described in example 2. Transfection efficiencies were higher than 95 % in all cases, as judged by β -galactosidase staining *in situ*. At the end of the chase period cells were lysed in SDS and aliquots of the cell lysates were analyzed by SDS-PAGE under reducing conditions. The amount of radioactivity associated with the fusion proteins was quantitated as described in example 2. Fig. 3 shows the fusion protein production (expressed as percentage of SFV μ SQL C-protein production) as a function of the number of C-protein derived amino acids in the fusion protein. The protein production from the longer constructs (containing more than 34 C-protein derived amino acid residues) reached a level of 85 % of the wild-type C-protein production. This is as high as that obtained from the SFVC-LacZ construct described in example 2. The shorter ones (containing 16, 19 and 27 C-protein derived amino acid residues, respectively) were produced at a considerably lower level. Thus, the results demonstrate that a 102 bases long portion of the 5' C-gene (b1) was mediating the increased expression.

In this example RNA transcription, transfection, labelling and SDS-PAGE were done as described in example 2. β -galactosidase staining of methanol fixed BHK-21 cells in tissue culture dishes was done as described by Sanes ⁽²³⁾.

Example 4

Expression enhancement by the 102 basis long C-gene segment is mediated through increased translation.

The fact that only a short stretch of the C-gene sequences was needed for optimal protein production makes it unlikely that the effect is mediated *in trans* (i.e. by the capsid polypeptide chain). A *cis* acting (i.e. RNA mediated) mechanism, which influences the efficiencies of either transcription or translation, would seem to be a more likely explanation. In order to

determine whether the C-gene segment affected the levels of genomic and subgenomic RNAs, the intracellular amounts of these two RNA species were measured in cells transfected with either "high" or "low" producer genomes. Transfected cells were cultured in the presence of Actinomycin D in order to suppress any residual host cell RNA synthesis. At 4 h post transfection ^{14}C -uridine was added and cells were incubated for 4 h. Total RNA was prepared from the labelled cells and subjected to electrophoresis through 1.0 % agarose gels under denaturing conditions. Gels were dried and the amount of radioactivity in different bands were quantitated as described in example 2. Fig. 4 shows an autoradiography of a dried gel: lane 1, SFV-mSQL; lane 2, the high producing SFVC'b7-LacZ; lane 3, the low producing pSFV3-LacZ; lane 4, mock transfection without RNA and lane 5, SFV1-Pr55^{gag}; which contains the coding sequence of the Gag-precursor of Human immunodeficiency virus type I. The subgenomic RNA of the SFV1-Pr55^{gag} genome is about 1500 bases shorter than that of SFV-mSQL and the SFVC-LacZ genomes, thus, facilitating the identification of these transcripts. Subgenomic RNAs were found to be synthesized at comparable levels in all transfections. Quantitations indicated that the levels of synthesis varied at the most by a factor of two and did not correlate with the level of protein production. Nor were there any significant differences detected in the levels of the genomic RNA. Thus, neither the replication nor the transcription of the various genomes were effected. Therefore, it is concluded that the increased protein production is a consequence of increased translational efficiency.

In this example ^{14}C -uridine labelling was done as follows: Transfected cells were directly plated and incubated for 2 h at 37°C. Then 1.0 $\mu\text{g/ml}$ of Actinomycin D was added to the medium and the incuba-

tion continued for 2 h. At 4 h post transfection the medium was removed, 75 kBq of ^{14}C -uridine (2.1 GBq/mmol; NEC-167, Dupont) was added in 1.5 ml complete BHK medium and the cells were incubated for 4 h at 37°. The labelling medium was removed and the cells lysed by addition of 1.0 ml of a phenol based nucleic acid extraction cocktail (Trizol TM; Gibco).

RNA purification and denaturing agarose gel electrophoresis was done as follows: Total RNA from ^{14}C -uridine labelled cells was purified by Trizol TM system as described by the manufacturer (Gibco) and dissolved in 20 μl RNase free H_2O . Aliquots were prepared for formaldehyde agarose gel electrophoresis and the RNA was separated in a 1.0 % Seaplaque agarose (FMC Bioproducts, USA) gel as described (Sambrook 1989). After electrophoresis the gel was immersed in 10 % trichloroacetic acid, incubated at a room temperature for 30 min and dried. The radio-labelled RNA bands were visualized by autoradiography and quantitated as described in example 2.

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CLAIMS

1. A self-replicative and transcription competent recombinant alphavirus RNA molecule comprising at least part of an alphavirus RNA genome and heterologous RNA encoding a substance having biological activity located downstream of an alphavirus base sequence having translation enhancing activity, said translation enhancing base sequence being comprised of a complete alphavirus capsid gene or a 5' portion of said gene.
2. The recombinant RNA molecule of claim 1, wherein the heterologous RNA is inserted into the alphavirus subgenome region after the said translation enhancing base sequence, which RNA molecule, after being introduced into cells, is replicated using alphavirus replicase encoded by the 5' region of the molecule and transcribed by the alphavirus subgenome promoter into mRNA from which the heterologous gene product is expressed in the cells as a fusion product with the complete capsid protein or with an aminoterminal portion of the capsid protein encoded by a 5' portion of said gene.
3. The recombinant RNA molecule of claim 1 or 2, wherein the alphavirus is Semliki Forest virus.
4. The recombinant RNA molecule of claim 3, wherein the translation enhancing base sequence is comprised of the first 81, and preferably the first 102 bases of the capsid gene having the sequence

ATGAATTACA	TCCCTACGCA	AACGTTTTAC
GGCCGCCCGGT	GGCGCCCGCG	CCCGGCCGCC
CGTCCTTGGC	CGTTGCAGGC	CACTCCGGTG
GCTCCCGTCG	TC.	
5. The recombinant RNA molecule of claim 2, 3 or 4, the heterologous RNA of which being expressed as the said fusion product, which is cleaved at its fusion site by the autolytic protease activity of the capsid pro-

tein.

6. The recombinant RNA molecule of any of claims 1-5, wherein the heterologous RNA sequence encodes a protein or a polypeptide comprising an antigenic determinant, said protein or polypeptide being immunogenic or antigenic.
7. The recombinant RNA molecule of any of claims 1-5, wherein the heterologous RNA encodes a protein or a polypeptide having therapeutic activity.
8. A DNA molecule, which comprises DNA sequences complementary to the recombinant RNA molecule of any of claims 1-7.
9. The DNA molecule of claim 8, which further comprises a DNA sequence encoding a promoter for transcribing the recombinant RNA molecule in cells, and further DNA sequences encoding traits required for plasmid growth in *E. coli*.
10. The DNA molecule of claim 9, wherein the promoter is SP6.
11. A transformed cell comprising a recombinant RNA or DNA molecule of any preceding claim.
12. The cell of claim 11 which is an animal or human cell.
13. A cell line comprising cells of claim 11 or 12, which are stably transformed with the said RNA or DNA molecule.
14. A method for expressing a heterologous RNA sequence encoding a biologically active substance, comprising transformation of cells with recombinant RNA of any of claims 1-7 or recombinant DNA of claim 8, 9 or 10, and subsequent growth of the transformed cells under conditions which allow for expression of the heterologous RNA sequence.
15. The method of claim 14, wherein transformation is achieved by transfection.
16. The method of claim 14 or 15, wherein the RNA encodes a protein, polypeptide or peptide having therapeutic activity or prophylactic, such as

immunogenic or antigenic, activity.

17. The method of any of claims 14-16, wherein the expression product is isolated from the cells and optionally purified.
18. A method for expressing a heterologous RNA sequence encoding a biologically active substance, comprising transformation, such as transfection, of cells *in vivo* in an animal or human individual with recombinant RNA of any of claims 1-7 or recombinant DNA of claim 8, 9 or 10.
19. A method to produce a recombinant alphavirus comprising a recombinant alphavirus RNA genome surrounded by a wild-type alphavirus coat, the said RNA genome comprising the recombinant RNA molecule of any of claims 1-7, by co-transformation, such as cotransfection, of cells with the said recombinant RNA genome and a helper RNA containing expression capacity of alphavirus structural proteins and comprising coding sequences for the alphavirus structural proteins, *cis* acting replication signals but no encapsidation signals, incubating the cells and collecting medium containing infectious recombinant alphavirus particles.
20. A transformed cell comprising a recombinant RNA molecule of any of claims 1-7 and a helper RNA containing expression capacity of alphavirus structural proteins and comprising coding sequences for the alphavirus structural proteins, *cis* acting replication signals but no encapsidation signals, which cell can produce infectious recombinant alphavirus particles comprising the said recombinant RNA surrounded by a wild-type alphavirus coat.
21. An infectious recombinant alphavirus particle comprising a genome consisting of a recombinant RNA molecule of any of claims 1-7 surrounded by a wild-type alphavirus coat.
22. A method for expressing a heterologous RNA sequence encoding a biologically active substance comprising

- infection of cells in cell culture or in an animal or human individual with recombinant alphavirus particles of claim 21.
23. The method of claim 22, wherein the RNA encodes a protein, polypeptide or peptide having therapeutic activity or prophylactic, such as immunogenic or antigenic, activity.
 24. The method of any of claims 22 or 23, wherein expression is achieved in cell culture and the expression product is isolated from the cells and optionally purified.
 25. The method of claim 22 or 23, wherein expression is achieved in an animal and the expression product is recovered by collection of a body fluid from said animal.
 26. A method for expressing a heterologous RNA sequence encoding a biologically active substance in an animal or human individual to achieve a therapeutic or prophylactic activity in said individual, said method comprising administration of cells according to claim 20 to said individual.
 27. The method of claim 26, wherein the heterologous RNA sequence encodes an immunogen or antigen giving rise to immunization of the individual wherein it is expressed.
 28. A pharmaceutical preparation comprising the recombinant RNA of any of claims 1-7 in a physiologically administrable form.
 29. The preparation of claim 28 comprising transformed cells according to claim 20.
 30. The preparation of claim 28 comprising infectious particles according to claim 21.
 31. Use of the recombinant RNA of any of claims 1-7 in the preparation of a pharmaceutical preparation according to claim 28 for use to achieve expression of the recombinant RNA in an animal or human individual.

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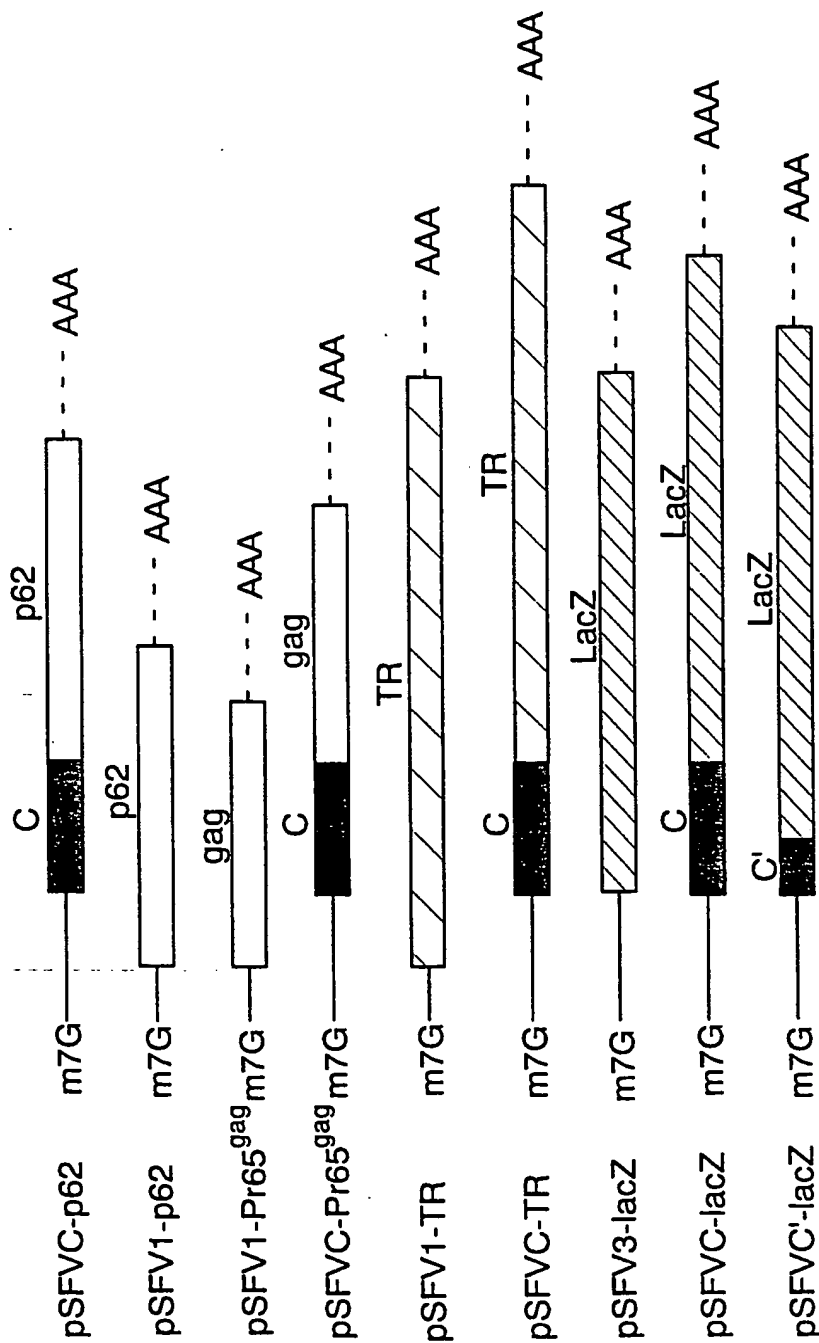


FIG.1

pSFVC'-lacZ	
Variant part	Length (amino acid residues)
c17	16
a2	19
b26	27
b1	34
b7	48
b11	72
b21	81
b22	103

2/3

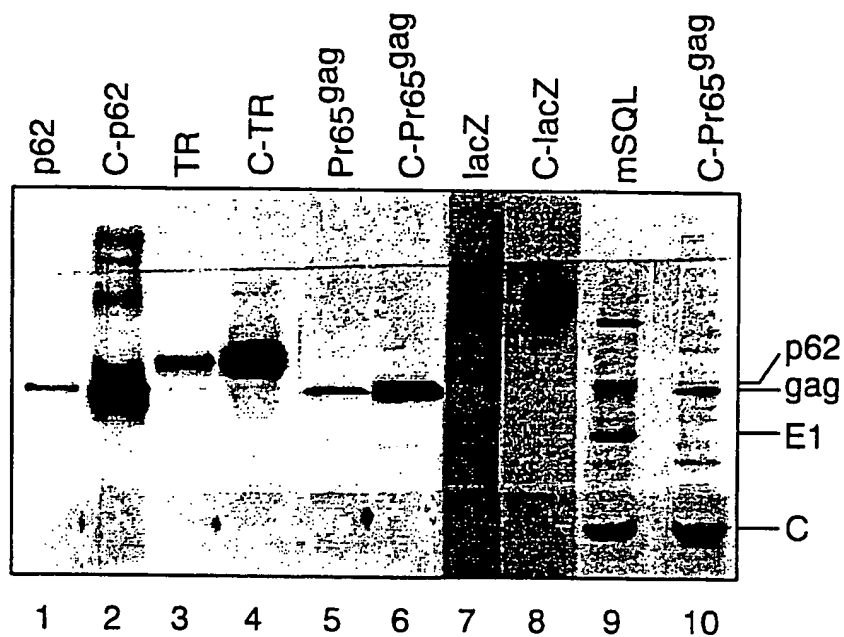


FIG.2

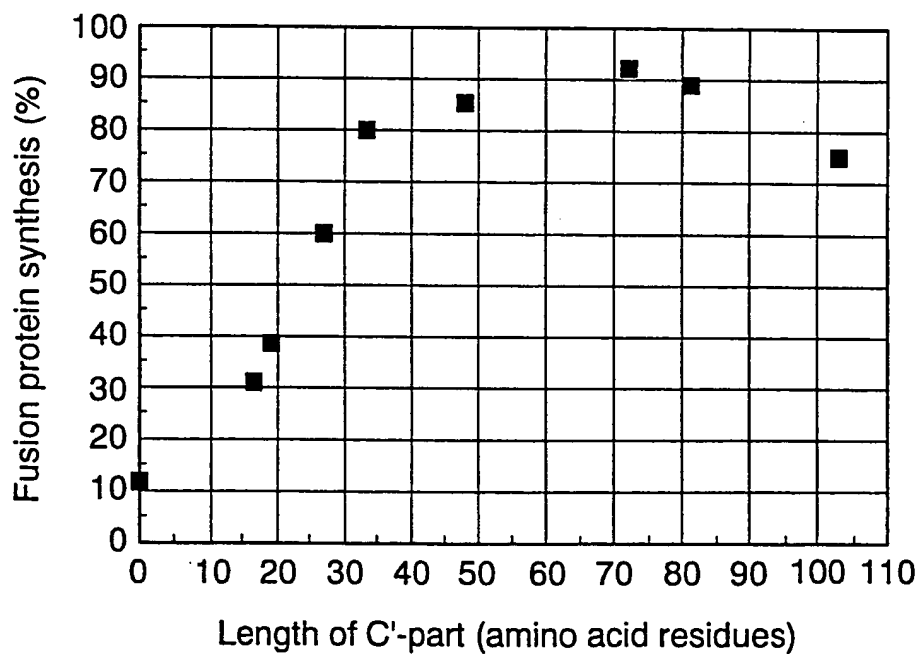


FIG.3

SUBSTITUTE SHEET

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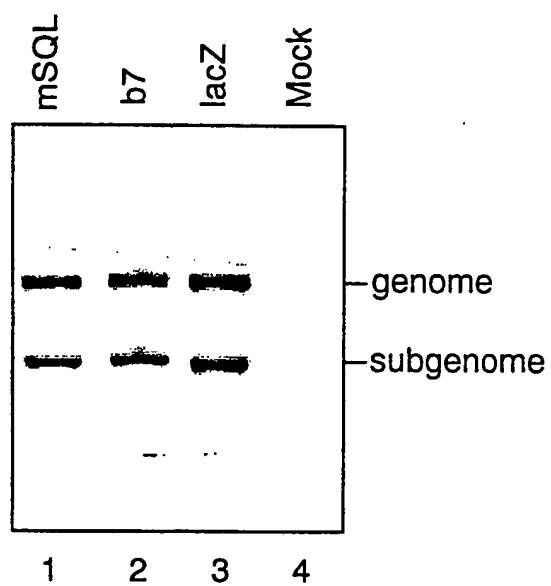


FIG.4

INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 95/00558

A. CLASSIFICATION OF SUBJECT MATTER

IPC6: C12N 15/86, C12N 7/01

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC6: C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	BIO/TECHNOLOGY, Volym 9, December 1991, Peter Liljeström et al, "A New Generation of Animal Cell Expression Vectors Based on the Semliki Forest Virus Replicon" sida 1356 - sida 1361 --	1-21,28-31
A	WO 9210578 A1 (BIOPTION AB), 25 June 1992 (25.06.92) --	1-21,28-31
A	US 5217879 A (HENRY V. HUANG ET AL), 8 June 1993 (08.06.93) --	1-21,28-31

☒ Further documents are listed in the continuation of Box C.☒ See patent family annex.

* Special categories of cited documents:

- "A" document defining the general state of the art which is not considered to be of particular relevance
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Date of the actual completion of the international search

25 August 1995

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29 -08- 1995

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 95/00558

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	SCIENCE, Volym 243, 1989, Cheng Xiong et al, "Sindbis Virus: An Efficient, Broad Host Range Vector for Gene Expression in Animal Cells" sida 1188 - sida 1191 -- -----	1-21,28-31

INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 95/00558

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 22-27
because they relate to subject matter not required to be searched by this Authority, namely:
PCT Rule 39.1 (iv): Methods for treatment of the human or animal body by therapy.
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT
Information on patent family members

31/07/95

International application No.
PCT/SE 95/00558

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A1- 9210578	25/06/92	AU-B- 656545 AU-A- 9078791 EP-A- 0561890 FI-D- 932678 JP-T- 6504198	09/02/95 08/07/92 29/09/93 00/00/00 19/05/94
US-A- 5217879	08/06/93	NONE	

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